

Prospective pilot study for evaluation of high-flow oxygen therapy in dyspnoeic dogs: the HOT-DOG study

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1 **Prospective pilot study for evaluation of High-flow Oxygen Therapy in**
2 **dyspneic dogs: the HOT-DOG study**

3

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18

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20

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23

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26

27 **Abstract**

28 **Objectives:** To describe the use, effectiveness and tolerance of high-flow oxygen therapy in
29 dyspneic dog.

30 **Methods:** Prospectively, dogs in acute respiratory distress admitted to the ICU between January
31 and May 2018 that failed to respond to nasal oxygen therapy and medical stabilization after 30
32 minutes were transitioned to high-flow oxygen therapy and included in the study. High-flow
33 oxygen therapy, delivering an inspired oxygen fraction of 100%, was carried out using an
34 air/oxygen blender, active humidifier, single heated tube, and specific nasal cannula. Respiratory
35 rate (RR), pulse oxymetry (SpO₂), heart rate (HR), and tolerance score were assessed every 15
36 minutes from T₀ (under nasal oxygen) to 1 hour (T₆₀), and PaO₂ and PaCO₂ at T₀ and T₆₀.
37 Complications were recorded for each dog.

38 **Results:** Eleven dogs were included. At T₆₀, PaO₂, flow rate, and SpO₂ were significantly greater
39 than at T₀ (171 ±123 mmHg *vs* 73 ±24 mmHg; P=0.015; 18 ±12 L minute⁻¹ *vs* 3.2 ±2.0 L minute⁻¹,
40 P<0.01; 97.7 ±2.3% *vs* 91.6 ±7.2%, P=0.03, respectively). There was no significant difference in
41 PaCO₂, RR, HR between T₀ and T₆₀. Tolerance score was excellent, and no complication
42 occurred.

43 **Clinical significance:** This study established that high-flow oxygen therapy improves markers of
44 oxygenation in dyspneic dogs and confirms that this technique deliver effective oxygen with
45 comfort and minimal complications in this population.

46

47

48 **Introduction**

49 Supplemental oxygen administration is the first supportive measure provided to dyspneic
50 dogs upon admission to an emergency facility. In veterinary medicine, oxygen therapy is mostly
51 delivered by non-invasive techniques such as flow-by, nasal prongs, or oxygen cages. These
52 methods are known as conventional oxygen therapy (COT) and can achieve variable fractions of
53 inspired oxygen (FIO₂) ranging from 21 to 70% (Sumner & Rozanski 2013). When patients
54 remain hypoxemic despite COT, clinicians may choose among more advanced oxygen delivery
55 methods. One option is mechanical ventilation, which however requires prolonged general
56 anaesthesia, continuous advanced care and is associated with frequent complications, high costs,
57 and, depending on the underlying disease, a guarded prognosis - often leading the owner to a
58 decision of euthanasia (Hopper *et al.* 2007, Mueller 2007). In human medicine, non-invasive
59 ventilation (NIV) is a popular method to avoid intubation. Continuous positive airway pressure
60 (CPAP) is a NIV mode that has been validated in dogs (Briganti *et al.* 2010, Staffieri *et al.* 2014,
61 Meira *et al.* 2018). Continuous positive airway pressure is effective to improve arterial partial
62 pressure of oxygen (PaO₂) in companion animals when used with devices such as masks or
63 helmets (Briganti *et al.* 2010, Staffieri *et al.* 2014, Meira *et al.* 2018) and can be used in dyspneic
64 animals.

65 In human medicine, a non-invasive oxygen delivery technique known as high-flow oxygen
66 therapy (HOT) has emerged as an alternative to COT (Helviz *et al.* 2018). This system allows for
67 delivery of heated and fully humidified gas with high flow rates of up to 60 L minute⁻¹ through a
68 specific high-flow nasal cannula, with a FIO₂ ranging from 21% to 100% (Frat *et al.* 2015). Such
69 high flow rates are not achievable with COT because of technical limitations (flowmeters rates,
70 diameter of the nasal prongs). Furthermore, administration of a partially humidified, cold gas
71 could cause patient discomfort, desiccation of the nasal mucosa, airway constriction, impairment

72 of the mucociliary functions, and increased risk of infection if delivered at similar rates (Kallstrom
73 2002, Dunphy *et al.* 2004, Kilgour *et al.* 2004, Kopelman & Holbert 2003).

74 High-flow oxygen therapy uses an air-oxygen blender connected to a flow meter, an active
75 humidifier and heater, a heated breathing circuit, and a specific nasal cannula (Nishimura 2016)
76 (Figure 1). Warm and adequately humidified gas delivered at high flow rates has apparent
77 beneficial physiological effects that are increasingly recognized in humans. These include: good
78 tolerance (Sztrymf *et al.* 2012) and comfort (Roca *et al.* 2010, Frat *et al.* 2015), decreased
79 mismatch between the oxygen flow and the patient's inspiratory flow (Sztrymf *et al.* 2012),
80 allowing precise setting by the clinician; generation of a low level of positive airway pressure
81 (Groves & Tobin 2007, Parke *et al.* 2011), decreased anatomical dead space by washing the
82 expired volume of carbon dioxide from the airway and replacing it with oxygen-enriched gas
83 (Helviz *et al.* 2018, Hernández *et al.* 2017, Millar *et al.* 2014), and decreased ventilatory drive and
84 work of breathing in obstructive human patients (Di Mussi *et al.* 2018), allowing the use of HOT
85 in hypoxemic and hypercapnic respiratory failure.

86 High flow oxygen therapy has recently been studied in veterinary medicine. It has been
87 shown to be a safe and effective method for oxygen delivery in sedated and awake healthy dogs
88 (Daly *et al.* 2017; Jagodich *et al.* 2019), and a preliminary retrospective study described its
89 successful use in six hypoxemic dogs (Keir *et al.* 2016). No prospective study reporting the use of
90 HOT in dyspneic dogs has been conducted. The objective of this pilot study was to prospectively
91 describe the use, tolerance and potential complications of HOT in dyspneic dogs.

92

93 **Materials and Methods**

94 **Ethical statement**

95 The study protocol was approved by the VetAgro Sup Ethics committee (number 1730).

96

97 **Animals**

98 This prospective pilot study was conducted between January and May 2018 in the
99 emergency and critical care unit (SIAMU, VetAgro Sup) on client-owned dogs admitted for acute
100 respiratory distress. Respiratory distress at admission was defined as a respiratory rate (RR) above
101 40 breaths minute⁻¹ associated with an abnormal respiratory pattern or laboured breathing at rest.

102

103 **Inclusion**

104 After admission, all respiratory distressed patients received 100% oxygen delivered via a
105 single nasal cannula at 150 mL kg⁻¹ minute⁻¹ (Dunphy *et al.* 2002, Mazzaferro 2015), using a
106 standard wall water humidifier (COT). Standard therapy for stabilization of the respiratory distress
107 was administered based on the most likely diagnosis, at the discretion of the attending clinician,
108 independently from the study. After 30 minutes of medical stabilization, dogs non-responsive to
109 nasal oxygen therapy were transitioned to HOT and prospectively enrolled in the study. Dogs non-
110 responsive to nasal oxygen therapy (inclusion criteria) were defined as persisting respiratory
111 distress as previously defined or a SpO₂<95% under nasal oxygen at the end of the 30-minute
112 stabilization period. Owner consent was obtained prior to transition from nasal oxygen therapy to
113 HOT.

114

115 **Exclusion criteria**

116 The exclusion criteria were a body weight below 2 kg, adequate response to COT defined
117 as respiratory distress improvement and SpO₂>95%, or criteria for immediate intubation, defined
118 as excessive respiratory effort with impending respiratory fatigue or failure, severe hypoxemia
119 despite oxygen therapy (SpO₂<90% or PaO₂<60 mmHg), or a severe hypercapnia (arterial partial
120 pressure of carbon dioxide (PaCO₂)>60 mmHg).

121 Early termination criteria were failure to tolerate HOT (defined by a tolerance score (TS) equal to
122 4, Table 1) and fulfilment intubation criteria (as defined above). Dogs with a do-not-intubate order
123 remained on HOT despite being intubation candidates.

124

125 **High-flow oxygen technique description**

126 High flow oxygen therapy was delivered by a commercially available unit connected to a
127 specific nasal cannula interface (Optiflow®, Fisher&Paykel Healthcare). The system is composed
128 of an air-oxygen blender connected to oxygen wall source, a flow meter, an active humidifier and
129 heater, a heated breathing circuit, and a specific nasal cannula (Figure 1). The nasal cannula is a
130 soft silicone bilateral nasal prong with tubing that connects to the heated breathing circuit (Figure
131 2). The system allows for administration of humidified and warmed gas with a FIO₂ between 21
132 and 100%. Continuous humidification was ensured by a water chamber connected to a sterile
133 water bag. Temperature was set at 37°C and administered to the patient via the heated breathing
134 tube.

135 The nasal cannula was chosen according to the size of the dog's nostrils, so that the
136 cannula diameter would not exceed 50% of the diameter of the nostril and therefore allowed
137 exhalation with minimal resistance (Daly *et al.* 2017). Seven different sizes were available (4
138 junior, 3 adult). Once the size of the nasal cannula was chosen, they were gently tightened behind
139 the neck, and eventually secured with tape and sutured in place next to each nare (Figure 2).

140 In order to avoid recruitment of oxygen from the surrounding air and assure the delivery of
141 the predetermined FIO₂, the oxygen flow rate was set to be equivalent to the dog's minute
142 ventilation (MV=RR*tidal volume) (Helvis *et al.* 2018). For homogeneity, tidal volume was set at
143 10 mL kg⁻¹ (Grimm *et al.* 2015). For example, a 30 kg dog with a RR of 80 breaths minute⁻¹ had a
144 flow rate with HOT of 10*30*80=24 L minute⁻¹ (compared to a flow rate of 4.5 L minute⁻¹ with
145 nasal oxygen therapy). The FIO₂ was set at 100% and the gas temperature at 37°C during the 60-

146 minute protocol. Once setting was ready, the patient was connected to the heated breathing tube.
147 At the end of the 60-minute protocol period, the FIO₂ was adjusted to the lowest level possible to
148 maintain SpO₂>95%.

149

150 **Data recording**

151 The RR, SpO₂, heart rate (HR), TS (Table 1), flow rate, need for additional sedation, type
152 and dose of sedative agent used and complications during HOT were recorded by the same
153 operator (LH). Each of these parameters was recorded right after the 30-minute stabilization
154 period (T₀), when dogs were still receiving nasal oxygen therapy. The oxygen delivery technique
155 was then changed from nasal oxygen therapy to HOT and the parameters were recorded
156 immediately at 15, 30, 45 and 60 minutes after T₀ (T₁₅, T₃₀, T₄₅, and T₆₀, respectively). A catheter
157 was inserted in the dorsal pedal artery to allow drawing of arterial blood samples and evaluate
158 PaO₂ and PCO₂ at T₀ (under nasal oxygen therapy) and T₆₀ (under HOT). Arterial blood gases
159 measurements were performed on fresh whole arterial blood collected in a heparinized syringe
160 according to the manufacturer's instructions using an on-site VetStat (IDEXX Laboratories Inc.).

161 An APPLE_{fast} scale (0-50 score, calculated from glucose, albumin, lactate, platelet count,
162 and mentation score) was used in each dog at inclusion to stratify illness severity by mortality risk
163 as previously described (Hayes *et al.* 2010).

164

165 **Outcomes**

166 The evolution of PaO₂, PaCO₂ and flow rate between T₀ and T₆₀, and evolution of RR,
167 tolerance score, SpO₂ and HR every 15 minutes during the 60-minute protocol were recorded. The
168 occurrence of complications due to the oxygen delivery technique during the protocol,
169 requirement for escalation to intubation within 24 hours after admission and in-hospital mortality
170 were also recorded.

171

172 **Statistical method**

173 Statistical analyses were carried out with JMP® version 13.1 (SAS institute). All
174 individual data were described with a spaghetti plot graphic. Data were tested for normal
175 distribution with the Shapiro-Wilk test. Continuous variables were compared with paired t-tests.
176 Data were expressed as mean \pm standard deviation (SD) with 95% confidence intervals (CI). A
177 value of $P < 0.05$ was considered as statistically significant.

178

179 **Results**

180 **Animals**

181 During the study period, 11 dogs were included in the high flow oxygen therapy protocol.
182 The breeds of dogs included were three Golden Retriever, two King Charles Spaniel, two
183 Dachshunds and one each of Australian Shepherd, German Wirehaired Pointer, Jack Russell
184 Terrier and Pomeranian. Four dogs were female neutered and seven were male neutered, with
185 mean \pm SD weights of 28.3 ± 12.3 kg (range, 4.8 to 43.0 kg) and ages 7.8 ± 3.8 years (range, 2 to 13
186 years).

187 Five dogs had a diagnosis of aspiration pneumonia, and one of each: cardiogenic
188 pulmonary oedema, non-cardiogenic oedema following cluster seizures, leptospirosis infection,
189 pulmonary haemorrhages following a car accident, pulmonary hypertension of unknown origin,
190 and pericardial effusion. The mean \pm SD APPLE_{fast} score was 27.0 ± 4.4

191

192 **Recorded data**

193 Dogs were receiving nasal oxygen therapy at T₀ and HOT at T₁₅, T₃₀, T₄₅ and T₆₀.

194

195 *PaO₂*

196 High-flow oxygen therapy allowed a significant increase in mean PaO₂ at T₆₀ (171 ±123
197 mmHg) compared to nasal oxygen therapy at T₀ (73 ±24 mmHg; P=0.015, 95% CI: 23 to 172
198 mmHg).

199 Individual data are presented in Figure 3. For every dog, PaO₂ increased after HOT
200 initiation. Of the 7 dogs with PaO₂<80 mmHg at COT, 5 had a resolution of their hypoxemia one
201 hour after HOT initiation.

202

203 *Flow rate (Figure 4)*

204 The mean oxygen flow rate was significantly greater with HOT (18 ±12 L minute⁻¹) than
205 in nasal oxygen technique (3.2 ±2.0 L minute⁻¹, P<0.01, 95% CI: 8.0 to 22.5 L minute⁻¹). Flow
206 rate with HOT ranged from 2 to 37 L minute⁻¹. Flow rate with nasal oxygen technique ranged
207 from 0.7 to 6.5 L minute⁻¹.

208

209 *Respiratory rate*

210 Individual data are presented in Figure 5. Six dogs had a decrease in RR between T₀ and
211 T₆₀. However, only 2 dogs reached our criteria of respiratory distress of less than 40 breaths
212 minute⁻¹ at T₆₀ despite the use of HOT.

213

214 *SpO₂, HR and PaCO₂*

215 The mean SpO₂ was significantly greater at T₆₀ (97.7 ±2.3%) than at T₀ (91.6 ±7.2%,
216 P=0.03, 95% CI: 0.7 to 11.5%). There was no significant difference at any time for HR or PaCO₂.

217 All individual data are showed in figure 6, 7, and 8 (supplemental material).

218

219 *Tolerance score*

220 The TS remained low during the entire protocol. Only one animal displayed a TS of 3
221 when setting the HOT device, requiring additional sedation that lowered its TS to 2 or 1
222 depending on the protocol time. Another dog displayed a TS of 2 at two occurrences but did not
223 need any additional sedation. Every other dog had a TS of 1 at every protocol time.

224

225 *Complications*

226 No complications due to the oxygen delivery technique occurred during the study. Nasal
227 cannulas were easy to fix and to use. No injuries occurred due to the nasal cannula.

228

229 *Intubation within 24 hours and mortality in hospitalisation*

230 Five of the 11 dogs (45%) developed a need for intubation within 24 hours after
231 admission. Due to financial reasons, only 3 of them (27%) were intubated. Six of 11 dogs (54%,
232 including the 5 that presented intubation criteria) died during hospitalization. Death were due to
233 deteriorating clinical state leading to cardiac arrest in 5 dogs and 1 euthanasia for prognostic
234 reason.

235

236 **Discussion**

237 This is the first prospective clinical trial evaluating the effectiveness and tolerance of HOT
238 in respiratory distress dogs. Our study shows the potential for HOT to be an efficient, safe and
239 well-tolerated alternative to COT with nasal oxygen therapy to deliver oxygen. This technique
240 allows for a significant increase in PaO₂ in dogs, with the PaO₂ more than doubling between nasal
241 oxygen therapy and HOT within only one hour. This finding is consistent with the human
242 (Sztrymf *et al.* 2012, Frat *et al.* 2015, Makdee *et al.* 2017), and the veterinary literature. Indeed,
243 Daly *et al.* (2017) and Jagodich *et al.* (2019) in healthy dogs, and Keir *et al.* (2016) in hypoxemic
244 dogs showed this PaO₂ improvement after HOT. With such an increase in PaO₂ during a short

245 period of time, HOT raised concern for oxygen toxicity if continued for a long time. For
246 homogeneity in the study design, the authors chose to keep the FIO₂ up to 100% during the 60-
247 minute protocol, but FIO₂ was decreased immediately at the end of the protocol to the lowest
248 required FIO₂ to reach a SpO₂>95%. As high-flow devices offer multi-configurable solutions for
249 flow and FIO₂, these two parameters can be changed easily to fit patient's demand. Once the
250 targeted oxygenation parameters are reached and plateaued, the FIO₂ should be decreased under
251 60% as soon as possible to avoid oxygen toxicity (Guenther 2019).

252 In the present study, flow rates with HOT ranged from 2 to 37 L minute⁻¹ and resulted in
253 an increase in PaO₂ in all cases (Figure 3). By delivering higher flow rates, HOT systems are less
254 prone to allow inhalation of room air during patient inspiration than traditional oxygenation
255 systems (Helvis *et al.* 2018). Different systems exist for high-flow delivery: Precision flow
256 (Vapotherm), Optiflow (Fisher&Paykel Healthcare), Airvo system (Fisher&Paykel Healthcare).
257 Each of them has specific characteristics and has to be used with their specific nasal cannula to
258 ensure appropriate flow rate, temperature, and FIO₂. In our study, we used the Optiflow system
259 which was easy to use and offer a multi-configurable solution for patients requiring HOT. This
260 system relies on an O₂/air blender, and FIO₂ of 100% is obtained with use of oxygen wall not
261 mixed with air. No internal system was available to ensure that the delivered FIO₂ at the nasal
262 cannula was at 100%. We were not able to measure the delivered FIO₂ in our study, and some
263 dogs were panting during measurements, so we could not ensure that we were delivering the pre-
264 set FIO₂. Moreover, a recent study with the same system showed that, when delivering a FIO₂ of
265 100%, the effective delivered FIO₂ increases with flow rate and varies between 72.2% and 95%
266 for flow rate from 0.4 L kg⁻¹ minute⁻¹ to 2.5 L kg⁻¹ minute⁻¹ (Jagodich *et al.* 2019). In our study,
267 flow rate ranged from 0.44 to 1.84 L kg⁻¹ minute⁻¹, so we can hypothesise that the effective FIO₂
268 did not reach 100%. Despite this limitation, our study confirms that HOT allows increase in PaO₂
269 in clinical settings.

270 In human medicine, several studies have showed a significant decrease of RR after
271 initiation of HOT (Roca *et al.* 2010, Sztrymf *et al.* 2011, Sztrymf *et al.* 2012, Makdee *et al.* 2017).
272 In our study, six dogs had a decrease in RR between T₀ and T₆₀, but with very variable evolution
273 among time (Figure 5). The dogs were included in this study because of respiratory distress with
274 failure to respond to nasal oxygen therapy, that could reasonably not resolve in one hour,
275 especially as our population mainly had principally respiratory distress secondary to aspiration
276 pneumonia. Finally, they were treated with HOT early in the course of their respiratory distress
277 (30 min after admission), meaning that other treatments, such as diuretics, antibiotics, etc., did not
278 have time to be effective. Mackdee *et al.* (2017) showed a significant decrease in RR after one
279 hour in the HOT group compared to COT group in human patients with cardiogenic pulmonary
280 oedema. Another study comparing RR between patients under nasal oxygen therapy or HOT in a
281 more homogeneous population would be of great interest to differentiate between effects of time
282 versus effects of treatment.

283 By enhancing the heating and humidification of nasal cavities, HOT has been shown to
284 improve patient comfort in human medicine (Roca *et al.* 2010, Boyer *et al.* 2011, Sztrymf *et al.*
285 2012). In one veterinary study in hypoxemic dogs, HOT intolerance requiring sedation was
286 noticed in 1/6 dogs (Keir *et al.* 2016). However, no TS was used in this study. Jagodich *et al.*
287 (2019), using a different TS associated with a respiratory score, showed that HOT tolerance was
288 inversely proportional to flow rate, and flow rate above 2.5 L minutes⁻¹ were not well tolerated. In
289 this study, the HOT administered with specific nasal cannula was easily tolerated and displayed
290 very low TS during all the study period. It would have been interesting to compare tolerance of
291 COT and HOT, but it was not the purpose of this study, and more studies on HOT tolerance are
292 therefore needed. Moreover, one study in human medicine showed that temperature seems to
293 significantly impact the comfort of dyspneic patients with high flows: for equal flow, patient

294 comfort was significantly higher at 31°C compared to 37°C (Mauri *et al.* 2018). The impact of
295 temperature was not evaluated in our pilot study and could be assessed in a larger population.

296 No clinically relevant complications related to the oxygen therapy technique were noted
297 during our study, which therefore supports previously published studies of HOT safety in dogs
298 (Keir *et al.* 2016, Daly *et al.* 2017, Jagodich *et al.* 2019). Jagodich *et al.* (2019) showed that at
299 flow rates above 2 L kg⁻¹ minute⁻¹, dogs became less tolerant. Our study showed that lower flow
300 rate allowed improvement of oxygen parameters with good tolerance. However, in the study of
301 Daly *et al.* (2017), one dog had radiographic evidence of gastric distension, and in the study of
302 Jagodish *et al.* (2019), 8 out of 8 dogs had aerophagia noted on radiographs. As our dogs were
303 dyspneic, we did not perform any abdominal radiographs to evaluate the incidence of gastric
304 distension. Nevertheless, we did not notice any abdominal distension during physical examination.
305 Moreover, the potential long-term complications could not be assessed with our study design.

306 Given the inclusion criteria of severely dyspneic dogs, the mortality rate in our population
307 was high. In the study of Keir *et al.* (2016), 3 out of 6 dogs died, of those, 2 died as a result of
308 worsening hypoxemia. Our study population included various primary pulmonary disease
309 processes, that have a different prognosis (Hopper *et al.* 2007). However, both in Keir *et al.*
310 (2016) and in our study, aspiration pneumonia was the main diagnosis. Larger population is
311 needed to compare influence on mortality rate of HOT.

312 Some limitations should be noted in the current study. The biggest limitations are the small
313 sample size and the absence of control group that limits interpretation of the results. Our study
314 was designed as a pilot study for a future randomised controlled study. Adding the result of a
315 control group, treated with standard medical stabilisation and nasal oxygen therapy would have
316 been of great interest to provide context for the magnitude of improvement in recorded parameters
317 (i.e. effects of time versus treatment type). However, the dogs included in our study were severely
318 dyspneic, and not adding HOT to their therapeutic plan could have raised ethical concerns.

319 Second, HOT has been showed to have several physiological effects allowing improvement of
320 PaO₂. In dogs, Jagodish *et al.* (2019) showed that HOT provides CPAP and predictable oxygen
321 support in healthy dogs. As our study was a clinical study, it was not possible to use more invasive
322 monitoring to differentiate between a positive effect on PaO₂ related to a therapeutic effect of the
323 HOT on respiratory mechanic or related to enrichment of inspired oxygen. Third, the long-term
324 implications of HOT were not evaluated, and our results (especially tolerance and complication)
325 should be validated in a longer study. Finally, the inclusion of patients with different respiratory
326 distress causes could have change the results, as some patients could benefit more or less from
327 HOT.

328

329 **Conclusion**

330 This study is the first in veterinary medicine to prospectively confirm that HOT could
331 deliver effective oxygenation and comfort with minimal complications in dyspneic dogs, and
332 gives practical information on HOT use in dogs.

333

334 **Bibliography**

335 Boyer, A., Vargas, F., Delacre, M., *et al.* (2011) Prognostic Impact of High-Flow Nasal Cannula
336 Oxygen Supply in an ICU Patient with Pulmonary Fibrosis Complicated by Acute Respiratory
337 Failure. *Intensive Care Medicine* 37, 558-559

338 Briganti, A., Melanie, P., Portela, D., *et al.* (2010) Continuous positive airway pressure
339 administered via face mask in tranquilized dogs. *Journal of Veterinary Emergency and Critical*
340 *Care* 20, 503-508

341 Brown, J.E., Bersenas, A.M.E., Mathews, K.A., *et al.* (2009) Noninvasive ventilation in cats.
342 *Journal of Veterinary Emergency and Critical Care* 19, 416-425

343 Daly, J.L., Guenther, C.L., Haggerty, J.M., *et al.* (2017) Evaluation of oxygen administration with
344 a high-flow nasal cannula to clinically normal dogs. *American Journal of Veterinary Research* 78,
345 624-630

346 Di mussi, R., Spadaro, S., Stripoli, T., Volta, C.A., Trerotoli, P., Pierucci, P., Staffieri, F., Bruno,
347 F., Camporota, L., Grasso, S. (2018) High-flow nasal cannula oxygen therapy decreases
348 postextubation neuroventilatory drive and work of breathing in patients with chronic obstructive
349 pulmonary disease. *Critical Care* 22, 180

350 Dunphy, E., Mann, F.A., Dodam, J., Branson, K., Wagner-Mann C., Johnson P., Brady M. (2002)
351 Comparison of unilateral versus bilateral nasal catheters for oxygen administration in dogs.
352 *Journal of Veterinary Emergency and Critical Care* 12, 245–251

353 Frat, J.P., Thille, A.W., Mercat, A., *et al.* (2015) High-flow oxygen through nasal cannula in acute
354 hypoxemic respiratory failure. *The New England Journal of Medicine* 372, 2185-2196

355 Guenther, C.L. (2019) Oxygen therapy In: Textbook of small animal emergency medicine. 1st
356 edn. Eds K. Drobatz, K. Hopper; E. Rosanski and D. Silverstein. Wiley-Blackwell, Hoboken. pp
357 1177-1182

358 Grimm, K., Lamont, L., Tranquilli, W., *et al.* (2015) Anesthesia equipment. In: Veterinary
359 anesthesia and analgesia. 5th ed. Ames, Iowa, Wiley Blackwell. pp 23-85

360 Groves, N., Tobin, A. (2007) High flow nasal oxygen generates positive airway pressure in adult
361 volunteers. *Australian Critical Care: Official Journal of the Confederation of Australian Critical*
362 *Care Nurses* 20, 126-131

363 Hayes, G., Mathews, K., Doig, G., *et al.* (2010) The acute patient physiologic and laboratory
364 evaluation (APPLE) score: a severity of illness stratification system for hospitalized dogs. *Journal*
365 *of Veterinary Internal Medicine* 24, 1034-1047

366 Helviz, Y., Einav, S. (2018) A Systematic Review of the High-flow Nasal Cannula for Adult
367 Patients. *Critical Care* 22, 71

368 Hernández, G., Roca, O., Colinas, L. (2017). High-flow nasal cannula support therapy: new
369 insights and improving performance. *Critical Care* 21, 62

370 Hopper, K., Haskins, S.C., Kass, P.H., *et al.* (2007) Indications, management, and outcome of
371 long-term positive-pressure ventilation in dogs and cats: 148 cases (1990-2001). *Journal of the*
372 *American Veterinary Medical Association* 230, 64-75

373 Jagodich, T., Bersenas, A., Bateman, S., Kerr, C. (2019) Comparison of high flow nasal cannula
374 oxygen administration to traditional nasal cannula oxygen therapy in healthy dogs. *Journal of*
375 *Veterinary Emergency and Critical Care* DOI: 10.1111/vec.12817

376 Kallstrom, T.J. (2002) AARC Clinical Practice Guideline: oxygen therapy for adults in the acute
377 care facility--2002 revision & update. *Respiratory Care* 47, 717-720

378 Keir, I., Daly, J., Haggerty, J., *et al.* (2016) Retrospective evaluation of the effect of high flow
379 oxygen therapy delivered by nasal cannula on PaO₂ in dogs with moderate-to-severe hypoxemia.
380 *Journal of Veterinary Emergency and Critical Care* 26, 598-602

381 Kilgour, E., Rankin, N., Ryan, S., *et al.* (2004) Mucociliary function deteriorates in the clinical
382 range of inspired air temperature and humidity. *Intensive Care Medicine* 30, 1491-1494

383 Kopelman, A.E., Holbert, D. (2003) Use of Oxygen Cannulas in Extremely Low Birthweight
384 Infants is Associated with Mucosal Trauma and Bleeding, and Possibly with Coagulase-negative
385 Staphylococcal Sepsis. *Journal of Perinatology* 23, 94-97

386 Makdee, O., Monsomboon, A., Surabenjawong, U., *et al.* (2017) High-flow nasal cannula versus
387 conventional oxygen therapy in emergency department patients with cardiogenic pulmonary
388 edema: A Randomized Controlled Trial. *Annals of Emergency Medicine* 70, 465-472

389 Mazzaferro, E.M. (2015) Oxygen therapy. In: Small animal critical care medicine. 2nd edn. Eds
390 K. Hopper and D. Silverstein. W.B. Saunders, Philadelphia. pp 77-80

391 Meira, C., Joerger, F.B., Kutter, A.P.N., Waldmann, A., Ringer, S.K., Böehm, S.H., Iff, S.,
392 Mosing, M. (2018) Comparison of three continuous positive airway pressure (CPAP) interfaces in
393 healthy Beagle dogs during medetomidine-propofol constant rate infusions. *Veterinary*
394 *Anaesthesia Analgesia* 45,145-157

395 Millar, J., Lutton, S., O'Connor, P. (2014) The use of high-flow nasal oxygen therapy in the
396 management of hypercarbic respiratory failure. *Therapeutic Advances in Respiratory Disease* 8,
397 63-64

398 Mueller, E.R., (2007) Suggested Strategies for Ventilatory Management of Veterinary Patients
399 with Acute Respiratory Distress Syndrome. *Journal of Veterinary Emergency and Critical Care*
400 11, 191-198

401 Parke, R.L., Eccleston, M.L., McGuinness, S.P. (2011) The effects of flow on airway pressure
402 during nasal high-flow oxygen therapy. *Respiratory Care* 56, 1151-1155

403 Staffieri, F., Crovace, A., De Monte, V. *et al.* (2014) Noninvasive continuous positive airway
404 pressure delivered using a pediatric helmet in dogs recovering from general anesthesia. *Journal of*
405 *Veterinary Emergency and Critical Care* 24, 578-585

406 Sumner, C., Rozanski, E. (2013). Management of respiratory emergencies in small animals. *The*
407 *Veterinary Clinics of North America Small Animal Practice* 43, 799-815

408 Sztrymf, B., Messika, J., Bertrand, F., *et al.* (2011) Beneficial effects of humidified high flow
409 nasal oxygen in critical care patients: a prospective pilot study. *Intensive Care Medicine* 37, 1780-
410 1796

411 Sztrymf, B., Messika, J., Mayot, T., *et al.* (2012). Impact of high-flow nasal cannula oxygen
412 therapy on intensive care unit patients with acute respiratory failure: A prospective observational
413 study. *Journal of Critical Care* 27, 324.e9-13

1	The patient is calm, comfortable; no agitation; no attempts to remove the nasal cannula
2	The patient tolerates the nasal cannula, but looks stressed and afraid; no attempts to remove the nasal cannula
3	The patient is agitated and tries to remove the nasal cannula; it is still possible to keep the nasal cannula on by gently restraining the patient
4	The patient does not tolerate the nasal cannula, is agitated, attempts to pull the nasal cannula; additional sedation is required to tolerate the oxygen delivery device

Table 1 : Oxygen delivery device tolerance score (adapted from Staffieri *et al.* 2014)

Figure 1: Schematic representation of a high-flow oxygen device and its components (Drawing Dr Mathieu Taroni).

Figure 2: Dog with the specific high-flow nasal cannula

Figure 3: Change in Arterial Partial Pressure of Oxygen (PaO_2) for individual patients. HOT: High-flow oxygen therapy. In figures 3, 4, 5 and supplemental material, the same dog is represented by the same colours.

Figure 4: Change in flow rates for individual patients. HOT: High-flow oxygen therapy.

Figure 5: Change in respiratory rates (RR) for individual patients. HOT: High-flow oxygen therapy. *Data from one dog are missing.*

Supplemental material

Figure 6: Change in pulse oxymetry (SpO_2) for individual patients. HOT: High-flow oxygen therapy. *Data from one dog are missing.*

Figure 7: Change in heart rates (HR) for individual patients. HOT: High-flow oxygen therapy. *Data from one dog are missing.*

Figure 8: Change in arterial partial pressure of CO₂ (PaCO₂) for individual patients. HOT: High-flow oxygen therapy.

Conflicts of interest statement

No conflicts of interest have been declared.